

Sources of omega-6 fatty acids do not alter the rumen degradation and transit of fibre from dairy cow diets

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KEY WORDS: dairy cows, neutral detergent ABSTRACT. The objective of this study was to evaluate the behaviour fibre, omega-6 PUFA, lipid sources, ruminal of fibre in the digestive tract on the basis of the passage kinetics of forage kinetics and concentrate particles in cows fed different omega-6 fatty-acid sources. The scientific hypothesis of this study was that omega-6 fatty acids do not interfere with the digestion of fibre in the diets of dairy cows. Five primiparous lactating Holstein cows were used in the experiment. The experimental diets were: control (C), ground soyabean (GS), cottonseed (CS), soyabean oil (SO), calcium salts of fatty acids (CSFA). The global mean estimates for the parameters of passage rate (γ) were 0.038 and 0.055 h⁻¹ for forage and Received: 10 April 2013 concentrate, respectively. The only significant effect with respect to the passage Revised: 27 September 2013 rate was a high negative correlation between the concentrate passage rate and Accepted: 5 November 2013 dry matter intake. There was less undegradable neutral detergent fibre (NDF) in treatments without added lipid. Dietary supplementation with lipid sources does not alter the kinetic parameters of roughage and concentrate particle passage or in vitro NDF degradation. Sources of omega-6 fatty acids do not ⁵Corresponding author: e-mail: alter the rumen degradation and transit of fibre. jeffersongandra@gmail.com

Introduction

The effects of unsaturated fatty acids on ruminal digestion may be variable and the amount of roughage used in the diet can be considered a fundamental factor (Ueda et al., 2003). Interactions between degradation rate and passage rate determine the extent of digestion in a compartment and the amount of potentially degradable material in the faeces. Depending of the type of fatty acid (omega-6 or omega-3) and how it gets into the rumen, free or partially protected, this effect can be maximized or minimized. From the standpoint of rumen carbohydrate fermentation, biohydrogenation is a favourable process because it reduces the potential negative effects of unsaturated fatty acids on rumen fermentation of fibre. The negative effects on microbial fibre digestion are the primary reason why large amounts of free vegetable oils cannot be fed to dairy cows (Jalč et al., 2006a).

Unsaturated fatty acids are toxic to many species of rumen bacteria, particularly those that are involved in fibre digestion. Also, because of the anaerobic environment of the rumen, there is an excess of hydrogen that the microbial population continually needs to eliminate (Jalč et al., 2006b).

The objective of this study was to evaluate the behaviour of fibre in the digestive tract by using roughage and concentrate particle transit kinetics, and to evaluate the dynamics of the *in vitro* neutral detergent fibre degradation of dairy cows receiving diets with different omega-6 fatty acid sources.

Material and methods

Experiment 1 (in vivo)

The experiment was conducted in the Dairy Cattle Teaching, Research and Extension Unit of the Animal Science Department of the Viçosa Federal University, Viçosa, MG.

Five lactating primiparous Holstein cows that at the beginning of the experiment were 99 ± 22 days in lactation, had 480.4 ± 2.8 kg of body weight and a daily milk production of 20.14 ± 2.23 kg, were used. The animals were housed in tie-stalls. The experimental design was a 5×5 Latin square. Each experimental period lasted 21 days, of which the first 14 were for adaptation of the animals to the diets and sampling.

Treatments consisted of different dietary lipid sources: 1. control (without lipid supplementation), 2. ground soyabean (GS), 3. cottonseed (CS), 4. soyabean oil (SO), 5. calcium salts of fatty acids (Megalac-E®) (CSFA). The ratio of forage:concentrate was 55:45 on a dry matter (DM) basis; maize silage was used as roughage. All diets were formulated to be isoproteinous according to the NRC (2001). The analysed concentration of omega-6 PUFA of feed was: ground soyabean (50.18%), cottonseed (48.76%), soyabean oil (51.92%), and calcium salts of fatty acids (E-Megalac®) (34.36%). Feed intake was controlled daily by weighing the refusals and keeping them between 5% and 10% of the total offered (Table 1).

Parameters of the particle passage kinetics were estimated using chromium-mordanted fibre for maize silage, prepared according to Udén et al. (1980), and lanthanum (La) fixed to the soyabean meal fibre, prepared according to Hartnell and Satter (1979). Marked feeds were the same in all treatments. The animals received 75 g of chromium-marked fibre and 50 g of lanthanum-marked fibre, orally, Table 1. Composition of the experimental diets on dry matter basis, $\% \mbox{ DM}$

Indiana	Diets				
Indices	С	GS	CS	SO	CSFA
Ingredients					
maize silage	55.0	55.0	55.0	55.0	55.0
maize grain	24.7	24.0	21.0	25.0	25.0
raw soyabean grain	-	12.0	-	-	-
soyabean meal	12.0	6.0	11.0	15.0	15.0
calcium salts	-	-	-	-	2.5
cottonseed	-	-	10.0	-	-
soyabean oil	-	-	-	2.00	_
urea/ammonium sulfate (9:1)	1.3	1.0	1.0	1.0	1.0
wheat meal	5.0	-	-	-	-
sodium bicarbonate/ magnesium oxide (2:1)	0.4	0.4	0.4	0.4	0.4
ADE vitamins	0.1	0.1	0.1	0.1	0.1
mineral mixture ¹	1.5	1.5	1.5	1.5	1.0
Chemical composition, g · kg ⁻¹					
dry matter ²	587	593	592	589	588
organic matter ³	936	9.6	9.6	934	933
crude protein ³	159	16.0	15.8	157	157
fat ³	33	54	5.0	51	53
neutral detergent fibre ³	380	376	395	365	365
Fatty acids. g ⋅100 g ⁻¹ of FA					
C14:0	0.34	0.39	0.35	0.34	0.35
C16:0	15.28	15.68	13.67	15.18	15.19
C18:0	3.11	3.19	3.04	3.12	3.10
C18:1 cis	12.33	12.47	9.86	12.56	12.70
C18:2 (n-6)	20.86	28.43	23.78	32.92	33.04
C18:3 (n-3)	2.93	2.09	1.98	2.88	2.87
other	2.74	2.83	2.67	2.71	2.88

C – control (without fat source addition); GS – ground soyabean; CS – cottonseed; SO – soyabean oil; CSFA – calcium salts of fatty acids (Megalac-E®). Dry matter (DM), organic matter (OM), crude protein (CP), ether extract (EE), neutral detergent fibre (NDF); ¹ Composition in 100 kg: kg: dicalcium phosphate 42.5, limestone 25, sodium chloride 21, potassium chloride 7.5, ammonium sulphate 2.5, zinc sulphate 1.25; g: copper sulphate 250, cobalt sulphate 15, sodium selenite 5; ² g · kg⁻¹ of natural matter; ³ g · kg⁻¹ of dry matter

at 6.30 h of the 17th day of each experimental period. Faecal samples were collected directly from the rectum at 0, 4, 8, 12, 16, 24, 30, 36, 48, 56, 72, 96 and 120 h after the administration of the markers. Samples were dried in a forced-air drying oven (60°C) and ground in a cutting mill to pass a 1 mm sieve.

Faecal samples used to quantify the kinetic parameters of forage transit were analysed for DM. Chromium levels were determined by atomic absorption spectrophotometry using a nitrous oxide/acetylene flame, a wave length of 398.8 nm, and a gap width of 0.2 nm. In order to obtain the solutions and the readings of the samples containing lanthanum-chelated fibre, a method adapted from Hartnell and Satter (1979) was used. In it, the analysis was done by dry process, in a sand bath (200°C), using 5 g of sample in 100 ml crucibles and 6 N hydrochloric acid for the digestion. The parameters of the transit kinetics were estimated by adjusting the gama-2 time-dependent model, described by Ellis et al. (1994), to the faecal excretion of the indicator from each experimental unit:

 $C_t = Z \times (t - \tau) \times \gamma \times \exp \left[-\gamma \times (t - \tau)\right]$ (1) where: C_t – faecal concentration of the indicator in time 't' (ppm); t – time after the indicator administration (h); γ – time-dependent rate parameter relating to fibrous particles ruminal flow (h⁻¹); Z – parameter without direct biological interpretation (ppm × h); τ – time elapsed between the administration and the appearance of the indicator in the faeces (h).

The mean rumen-reticulum retention times were estimated according to Ellis et al. (1994):

$$RRRT = \frac{2}{\gamma}$$
(2)

where: RRRT – mean rumen-reticulum retention time (h); γ – as previously defined.

Experiment 2 (*in vitro*)

In order to evaluate, in vitro, the dynamics of NDF degradation, maize silage aliquots (192.5 mg of DM) were conditioned in 50 ml glass vials. The concentrates from each of the previously described treatments were then conditioned in the same vials in order to maintain the roughage:concentrate ratio used in the in vivo assay (55:45), making a total of 350 mg of DM per vial. After that, 28 ml of a buffer solution with a pH of 6.8 previously adjusted by CO, injection (McDougall, 1949), were added. Vials were kept in an acclimatized room (39°C) in order to hydrate the samples. During the hydration process, ruminal fluid was collected from a heifer, cannulated in the rumen and kept nearby the incubation room. Ruminal fluid was collected, filtered through a triple layer of gauze, conditioned in a thermos recipient, and immediately transported to the incubation room. Seven milliliters of the ruminal inoculum were added per vial, and immediately after that the incubation environment was saturated with CO₂ and the vials were sealed. Vials were kept at 39°C under orbital shaking (40 rpm). Fermentation gases were removed every three hours with the aid of needles. Evaluations were done at 0, 3, 6, 9, 12, 24, 36, 48, 72 and 96 h of incubation. The incubation procedure was repeated four times, totaling four evaluations per incubation time per treatment. At the end of each incubation time, vials were removed from the acclimatized room and their content was vacuum filtered in filter crucibles (coarse porosity).

Crucibles were conditioned in polyethylene flasks (120 ml), to which 50 ml of neutral detergent were added. After sealing, flasks were autoclaved (105°C/1 h), in order to extract all neutral detergent soluble components (micro-extraction technique). After that, another vacuum filtration, followed by washing of the residue in hot water and acetone, was done. The residual NDF was obtained after drying the material in an unventilated stove (105°C/16 h).

The NDF residues in time, for each treatment, were subjected through the PROC NLIN of SAS to an adjustment by the non-linear logistic model:

$$Rt = pdNDF \times (1 + \lambda \times t) \times exp(-\lambda \times t) + uNDF \quad (3)$$

were: Rt – undegraded NDF residue in time 't' (%); pdNDF – potentially degradable NDF fract ion (%); uNDF – undegradable NDF fraction (%); λ – combined fractional rate of latency and degradation of the pdNDF (h⁻¹); t – time (h).

Because parameter λ represents latency and degradation rates together, the fractional degradation rate was estimated from λ using the gama-2 distribution properties (Ellis et al., 1994):

$$kd = 0.59635 \times \lambda \tag{4}$$

where: kd – fractional degradation rate of the pdFDN (h^{-1}).

The estimates of the discrete latency were obtained according to the derivations from Vieira et al. (1997):

$$L = \frac{R(0) - R(t_i) + t_i}{R'(t_i)}$$
(5)

where: L – discrete latency (h); R(0) – undegraded NDF residue in t – 0 (%); $R(t_i)$ – undegraded NDF residue obtained at the inflection point of the degradation curve (%); $R'(t_i)$ – derived of the degradation-adjusted curve to the inflection point (maximum degradation rate of the substrate) (h⁻¹); t_i – time equivalent to the inflection point of the degradation curve (h).

The t_i values were obtained according to Van Milgen et al. (1991):

$$t_i = \frac{1}{\lambda} \tag{6}$$

The effectively degraded fractions of the NDF were obtained through an adaptation of the Ørskov and McDonald (1979) suggestions, according to the equation:

$$EDF = \lim_{t \to \infty} \int_{0}^{t} f(-\frac{dRT}{dt}) dt$$
 (7)

where: EDF – effectively degraded fraction of the NDF (%); f – function relative to the movement of the solids in the ruminal environment.

In order to define the function above (7), a ruminal movement of solids of gamma-1 order was assumed, according to the equation:

$$f = \exp(-k \times t) \tag{8}$$

where: k – fractional rate of solids movement in the ruminal environment (h⁻¹), which was obtained by weighing the passage rates of roughage solids and concentrate from the *in vivo* assay as a function of the participation of each one in the total diet NDF.

Thus, the effectively degraded fraction was estimated as:

$$EDF = pdNDF \times \frac{\lambda^2}{\left(\lambda + k\right)^2}$$
(9)

Statistical analyses

The non-linear adjustments were done by the Gauss-Newton interactive algorithm in PROC NLIN of the SAS programme (Statistical Analysis System, version 9.2). Parameters γ and RRRT estimates were compared by variance analysis according to a 5 × 5 Latin square design, considering treatment and experimental period as fixed effects, and animal as a variable effect. Data were analysed with PROC MIXED of SAS. Pearson's linear correlation coefficients were estimated for transit kinetics and intake and production characteristics. Estimates were obtained in an adjusted form for the effects of animal and experimental period by the MANOVA routine, implemented in PROC GLM of SAS.

Results

Productive performance

The level of supplementation was insufficient to promote significant changes in the fat content of milk (%). The daily milk yield in kg per day was higher (P < 0.05) in animals supplemented with CSFA in relation to those supplemented with SO and C. There was no difference (P > 0.05) among treatments for DM and NDF intake. The fat intake was higher (P < 0.05), however, for diets

 Table 2. Productive performance of lactating dairy cows fed diets with different lipid sources

láona	Experimental diets						D	
Item	С	GS	CS	SO	CSFA	-SEIVI	Ρ	
Intake, kg per o	day							
DM	17.09	17.10	16.59	16.73	17.10	0.78	0.932	
NDF	6.04	5.78	6.07	5.71	5.16	0.26	0.066	
fat	0.43 ^b	0.73ª	0.72ª	0.71ª	0.71ª	0.04	0.001	
Yield, kg per da	ay							
milk	18.48 ^b	19.89 ^{ab}	19.08 ^{ab}	18.70 ^b	21.90ª	2.46	0.023	
FCM, 3.5%	19.29	18.23	19.84	19.54	21.21	2.42	0.422	
Percentage								
fat	3.78	3.19	3.77	3.74	3.32	0.22	0.187	

C – control (without fat source addition); GS – ground soyabean; CS – cottonseed; SO – soyabean oil; CSFA – calcium salts of fatty acids (Megalac-E®); SEM – standard error of the mean

supplemented with lipids compared with the control treatment, with no difference among the experimental diets due to the characteristics of the treatments (Table 2). The performance data were used in this article only to emphasize the Pearson correlation analysis (Table 4) that is discussed in a subsequent article.

Ruminal kinetics

The ruminal passage of solids from the roughage or the concentrate fractions did not differ among diets (P > 0.05). The global mean estimate of the time-dependent rate (γ) parameter was 0.038 and 0.055 h⁻¹, for roughage (chromium) and concentrate (lanthanum), respectively. Despite the numerical differences, the retention time in the rumen-reticulum also did not differ among diets (P > 0.05), and averaged 52.84 and 38.15 h for roughage and concentrate, respectively (Table 3).

 Table 3. Estimates of fibre transit kinetics parameters in lactating cows

 fed diets with different lipid sources

Indices	Treatments						
	С	GS	CS	SO	CSFA	- P	
Rougha	ge						
γ	0.041 ±0.002	0.037 ±0.002	0.036 ±0.002	0.038 ±0.002	0.040 ±0.002	0.363	
RRRT	48.8 ±2.99	56.1 ±2.99	56.3 ±2.99	53.5 ±2.99	49.5 ±2.99	0.250	
Concentrate							
γ	0.058 ±0.005	0.055 ±0.005	0.051 ±0.005	0.054 ±0.007	0.055 ±0.005	0.902	
RRRT	34.7 ±4.31	36.8 ±4.31	41.2 ±4.31	40.6 ±6.08	36.6 ±4.31	0.722	

 γ – time-dependent rate-parameter relative to the ruminal flow of fibrous particles (h⁻¹); RRRT – average rumen-reticulum retention time; C – control (without fat source addition); GS – ground soyabean; CS – cottonseed; SO – soyabean oil; CSFA – calcium salts of fatty acids (Megalac-E®)

Table 4. Pearson's linear correlation coefficients among variables: dry matter intake (DMI); neutral detergent fibre intake (NDFI), 3.5% fatcorrected milk production (CMP – kg per day); milk fat percentage (F – %); time-dependent rate-parameter relative to the ruminal flow of roughage particles (γ cr – h^{-1}) and time-dependent rate-parameter relative to the ruminal flow of concentrate particles (γ *la* – h^{-1})

Variable ^{1,2}	Variable	1,2			
	DMI	F	CMP	Y _{CR}	Y LA
	0.441	0.028	-0.419	-0.270	-0.370
NDFI	0.099	0.920	0.119	0.328	0.174
ПΜ		-0.026	-0.017	0.065	-0.561
DIVII	-	0.924	0.951	0.815	0.029
F			0.556	-0.033	-0.186
		-	0.031	0.905	0.507
CMP				0.048	-0.105
			-	0.864	0.708
					0.207
γ _{CR}				-	0.457

¹ subscribed values correspond to descriptive levels of probability for type I error associated to the hypothesis: h_0 : $\rho = 0$; $\rho \neq 1$. 2 The correlation coefficients were adjusted for effects of animal and experimental period

Diet CS had the lowest numerical passage rate value, although not showing a significant effect. Diets CS and GS also showed numerically greater retention times in the rumen. The kinetic values found with the use of calcium salts, in this study, were the closest to the control diet. Concerning the treatment without added lipid supplement, the passage rate values were 0.0583 h^{-1} and 0.0412 h^{-1} for concentrate and roughage, respectively (Table 3).

Generally, the roughage (γ_{cr}) and concentrate (γ_{la}) passage rates had similar non-significant associations in relation to the NDF intake and F variables. The only significant effect, regarding the passage rate, was a high negative correlation (P < 0.05) between the concentrate passage rate (γ_{la}) and the DMI (Table 4).

In vitro degradation of NDF

There were no differences (P > 0.05) in the potentially degradable fraction of NDF (pdNDF), although the values obtained for the supplemented diets (GS, CS and SO) were, respectively, 12.69%, 5.59% and 11.07% lower, compared with the unsupplemented diet, except the CSFA diet, which had a degradation 2.93% higher than the control (Table 5).

The effectively degraded fraction (EDF_t) also did not differ among diets (P > 0.05), although the numerical values of diets GS, CS and SO were, respectively, 6.41%, 8.64% and 7.49% lower compared with the control diet, and 2.93% higher than diet CSFA (Table 4). The common rate of latency and degradation was also considered similar for all diets. Diets without lipid supplementation and with calcium salts had lower NDF undegradable fractions (I) (38.47% and 36.61%, respectively) than the others. The values for treatments GS, CS and SO were, respectively 16.88%, 8.21% and 15.04% higher than C.

Table 5. Percentage estimates of the potentially degradable fraction of the NDF (b), indegradable fraction of the NDF (i), effectively degraded fraction in relation to the total diet NDF (EDF₁), and of the common fractional rate for latency and degradation ($\lambda - h^{-1}$) and of the discrete latency rate for degradation of the NDF (LAG – h) for degradation profiles adjusted for different treatments

Treatment	Parameter							
	b	i	λ	EDFt	LAG			
С	61.5 ± 3.12	38.5 ± 2.62	0.08 ± 0.009	46.1	3.2			
GS	53.7 ± 2.93	46.3 ± 2.38	0.09 ± 0.011	43.1	2.9			
CS	58.1 ± 3.18	41.9 ± 2.90	0.07 ± 0.008	42.1	3.9			
SO	54.7 ± 3.11	45.3 ± 2.58	0.09 ± 0.011	42.6	3.1			
CSFA	63.4 ± 3.14	36.6 ± 2.77	0.08 ± 0.008	46.9	3.4			
Р	0.115	0.030	0.540	-	-			

C – control (without fat source addition); GS – ground soyabean; CS – cottonseed; SO – soyabean oil; CSFA – calcium salts of fatty acids (Megalac-E®)

Discussion

Ruminal kinetics

The passage rate of feed in the rumen is a variable of great importance, for it determines the digesta flow through the digestive tract. Greater passage rates are observed in highly digestible diets, with high daily intakes, and which are provided with greater frequency per day.

Because the roughage used in the different treatments was of the same kind and on the same level, and concentrates had a homogenous grinding pattern, except for the cottonseed that was provided whole and thus had greater particle size, no differences in passage rates were expected due to particle size. The amount of cottonseed required to keep the diets within the desired fat levels did not influence the particle passage rate.

The retention time in the rumen, calculated by the reciprocal passage rate in the organ, is correlated with the feeding level of the animal, as higher intakes result in greater passage rates and, consequently, higher nutrient supplies to the animal. The DM intake was relatively constant, however, and remained within normal limits for the evaluated animal category and production level.

The greater the degree of unsaturation of the fatty acid chain, the greater may be their interference in the kinetics of fibre in the gastrointestinal tract due to the decreased rate of passage that may be affected by decreased digestibility resulting from a possible deleterious action of some sources of fatty acids in the rumen (Jenkins, 1993).

Higher forage levels in the diet increase the ruminal retention time due to the slower fibre degradation rate, and higher amounts of fibre in the intestines reduce the post-ruminal retention time by stimulating peristalsis. Although the treatments had similar fibre levels, the origin and behaviour of this fibre differed according to the fatty-acid source utilized, especially relating to the cottonseed treatment. This may affect the RRRT when higher levels of lipid supplements are provided.

Costa (2008), evaluating lipid sources (control, soyabean, soyabean with formaldehyde, soyabean oil and calcium salts), also observed no difference (P > 0.05) in passage rates and RRRT among diets for roughage and soyabean meal particles. The mean values were 0.052 and 0.064 h⁻¹, for maize silage (chromium) and soyabean meal (ytterbium), respectively. Maize silage RRRT was 38.68 h and soyabean meal RRRT was 25.3 h.

The results observed for the CS diet can be explained by the presence of the cottonseed lint, which decreases the passage rate, altering ruminal fill and stimulating chewing, which contributes to increased digestibility. The lower roughage passage rate of GS diet, compared with the other diets, can be explained by the high hemicellulose content and by the cell wall structural characteristics of the oilseeds, in which the paired epidermis cells contribute to this increase in the length of the time of permanence, reducing digestibility and diet energy. Therefore, it is suggested that the peculiar characteristics of oilseeds are taken into account when used in their integral form.

Cecava and Parker (1993), working with steers fed maize silage and concentrate at a proportion of 62:38, observed a concentrate particle (ytterbium marked) passage rate of 0.0686 h^{-1} , close to that received by Bürger et al. (2000) under similar conditions, of 0.0679 h^{-1} for soyabean meal and 0.0412 h^{-1} for chromium-marked maize silage, higher than the values reported in this study with lanthanum, but similar to that observed for chromium.

Campos et al. (2007) suggested that the ruminal passage rates for forages are around 0.0416 h⁻¹, close to the average of the treatment without lipid addition in the present study. In the same way, the discrepancies between the values of γ and RRRT, in particular, found between the present study and those in the literature, demonstrate the need to obtain more research data on ruminal kinetics, in order to predict the mean disappearance of feed in the rumen in a more adequate form.

All diets contained the same concentration of fibre or NDF and similar fat concentrations, except the control diet, the only factor that could influence the degradation and kinetics of fibre would be the unsaturation of fatty acids of the various sources used, due the toxicity of these sources to fibredegrading bacteria. The sources of omega-6 were not able, however, to affect the process of ruminal digestion, due to the low degree of unsaturation of these sources. This same result could be different if the supplementation was with sources of omega-3 fatty acids, such as fish oil and linseed, due to the high degree of unsaturation and the size of these fatty-acid chains.

In vitro degradation of the NDF

The effective degradation of the NDF, although not statistically different, indicates that omega-6 fatty acid utilization may affect fibre degradability when used in ruminant diets at levels higher than the ones in this study, and that lipid protection through the formation of calcium salts of fatty acids is efficient in decreasing this degradation. Firkins et al. (1991), evaluating the *in vitro* fibre degradation, observed that addition of soyabean oil at 10% did not affect NDF digestion. Possibly, the small particle size and large surface area of the cellulose used as the fibre source may have dispersed the fat enough to avoid inhibition of NDF digestion by the active surface properties of fatty acids, since according to other studies, supplementation in this level affects fibre degradation.

Supplementation of diets of dairy cows with sources of omega-6 fatty acids does not really affect NDF degradability because it alters the basal ruminal fermentation as evidenced by Cieślak et al. (2013). The same authors conclude emphatically that supplementation with sources of omega-6 fatty acids did not adversely alter ruminal fermentation and, therefore, so also do not alter the degradation of NDF, which can no longer be said to be sources of omega-3 fatty acids.

Similar results were also obtained by Szumacher-Strabel et al. (2009), who observed no change in the concentrations of acetate and propionate when sources of omega-6 fatty acids were inoculated in the rumen of dairy cows, also indicating that there is no interference in the digestibility of NDF in these cases.

The release of omega-6 fatty acids from oilseeds is slow, providing small amounts of lipids in the ruminal environment, which may result in rapid biohydrogenation, avoiding unsaturated fatty acid accumulation and impairment of the ruminal degradation of the fibre. Moreover, the amount will not be enough to adhere to feed particles, causing physical impediment to microorganisms and microbial enzymes.

Conclusions

Inclusions of sources of omega-6 fatty acids do not interfere with digestion and kinetics of fibre in dairy cows in mid-lactation. Further studies should be performed in order to determine the behaviour of the fibre when cows are supplemented with sources of omega-3, as well as to establish digestion parameters in dairy cows in early lactation and the transition period, when supplemented with omega-3 and omega-6 fatty acids.

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